

The Resokine Pathway Is Implicated in the Pathology of Interstitial Lung Disease

L.A. Nangle^{1*}, Y. Tong^{2,3}, E. Mertsching¹, S.P. Crampton¹, R.A. Adams¹, K.P. Chiang¹, K.M. Ogilvie¹, P. Schimmel⁴, J.C. McKew¹, D. King¹, J.D. Mendlein¹

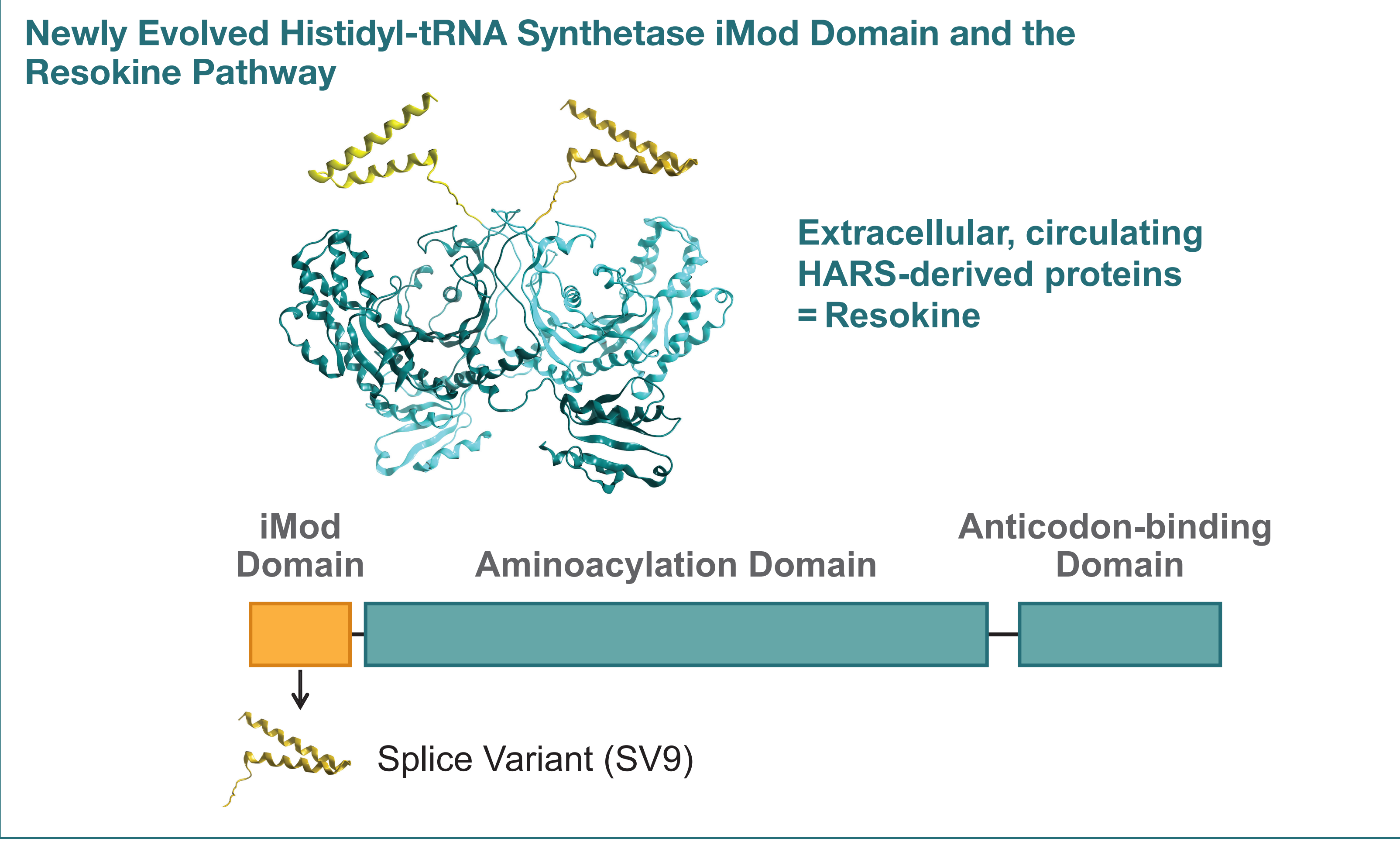
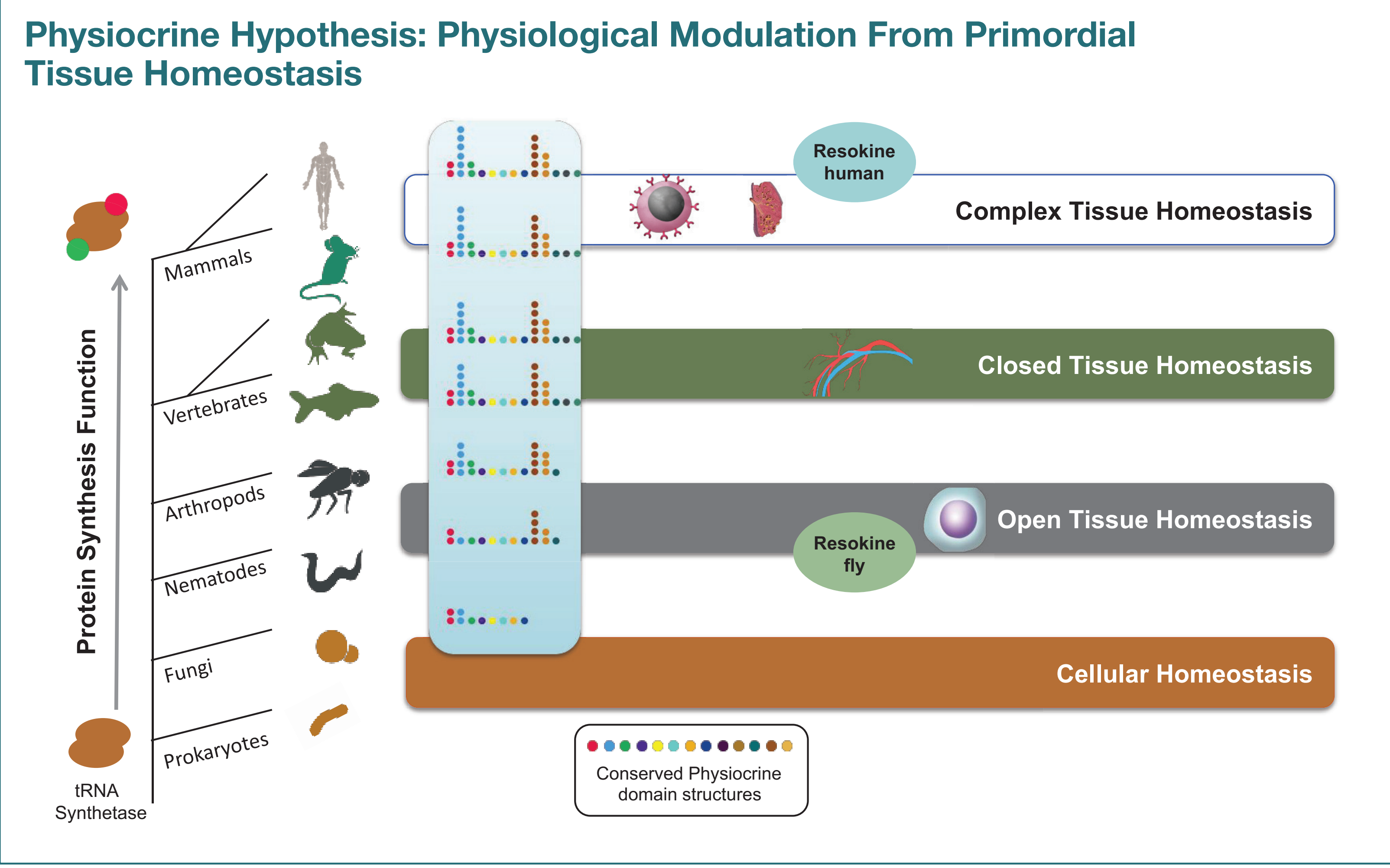
¹aTyr Pharma, San Diego, CA, USA; ²Pangu Biopharma, Hong Kong, China; ³IAS HKUST – Scripps R&D Laboratory, Institute for Advanced Study, Hong Kong, China; ⁴The Scripps Laboratories for tRNA Synthetase Research, The Scripps Research Institute, La Jolla, CA, USA

*Corresponding and presenting author: L.A. Nangle, lnangle@atyrpharma.com

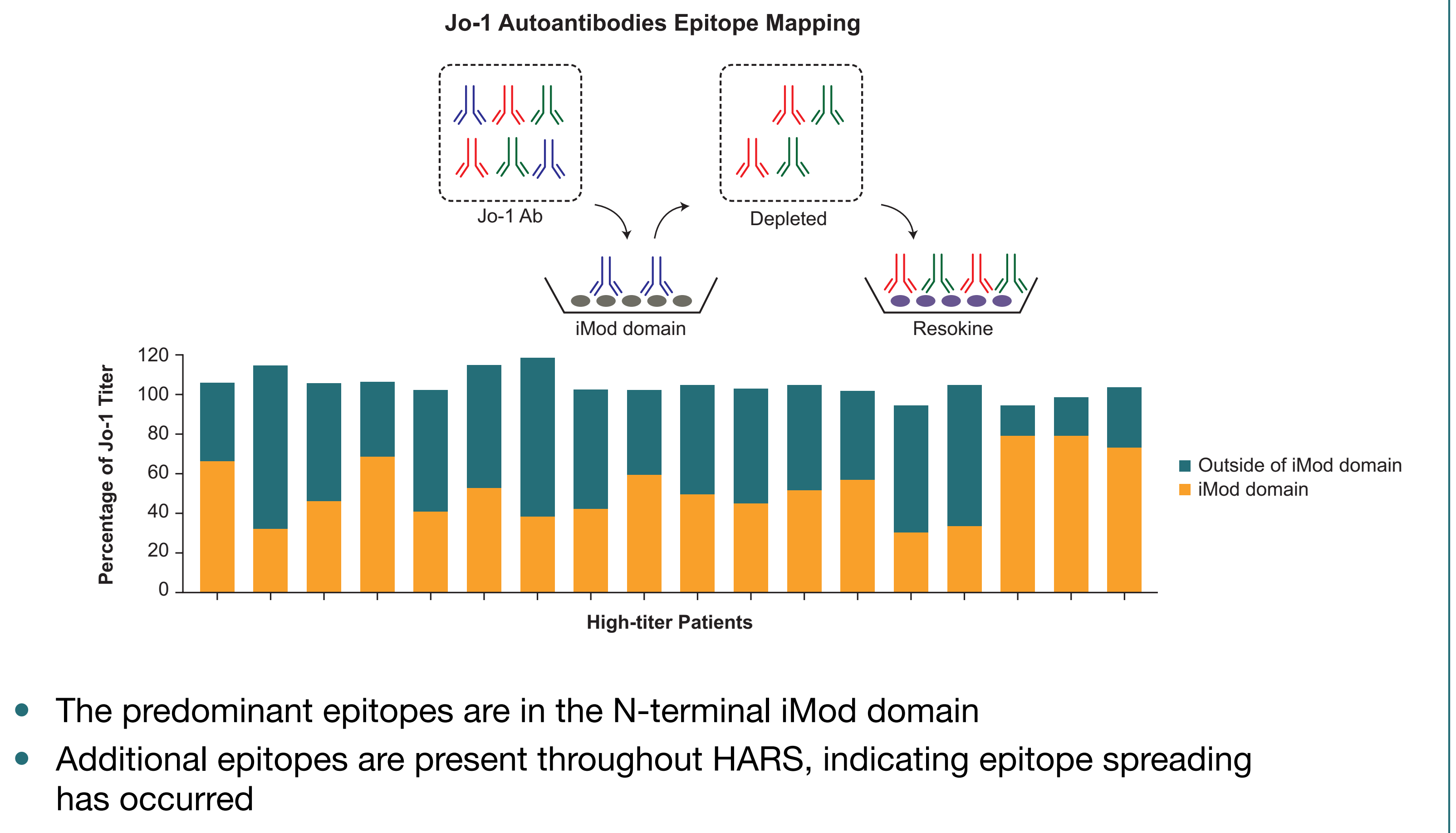
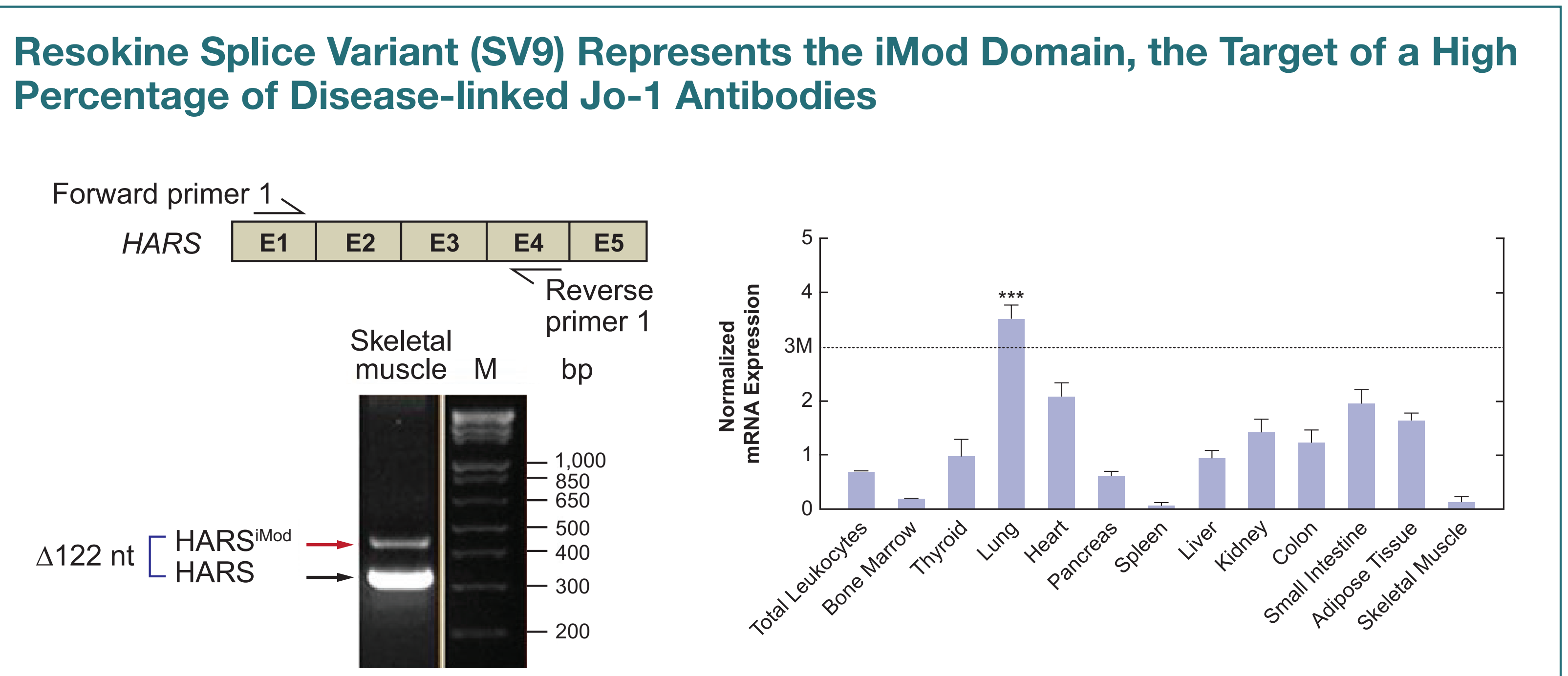
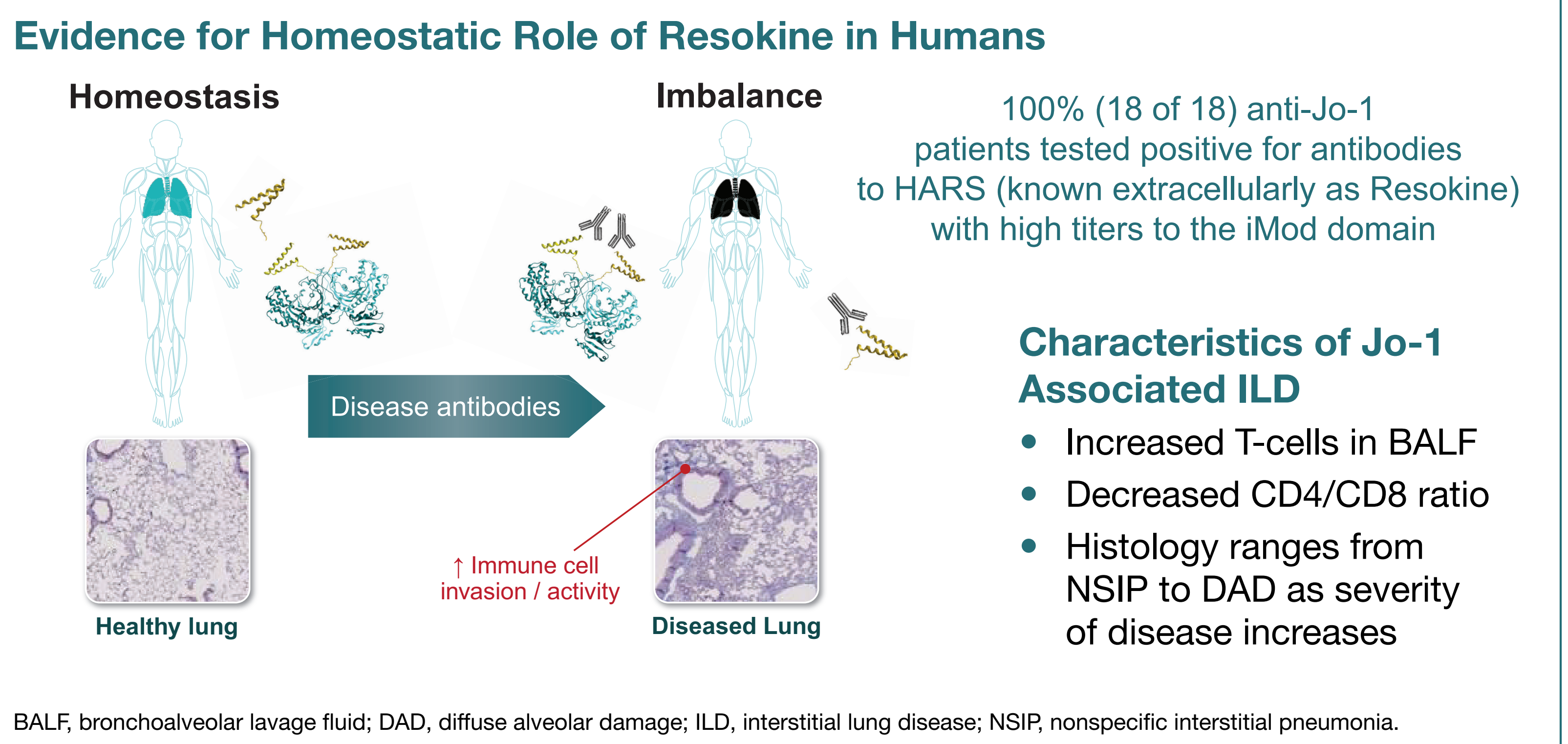
Rationale

- The histidyl-tRNA synthetase (HARS) protein has long been identified as the sole target of Jo-1 autoantibodies (Jo-1 Abs), which are, in many cases, accompanied by interstitial lung disease (ILD). Extracellular HARS proteins (Resokine) are found in circulation at physiologically relevant levels in healthy individuals (~200 pM) and are greatly reduced in patients with Jo-1 Abs, leading us to investigate the source of Resokine and explore the possibility that insufficiency plays a role in the pathology of inflammatory myopathies and ILD associated with Jo-1 Abs.

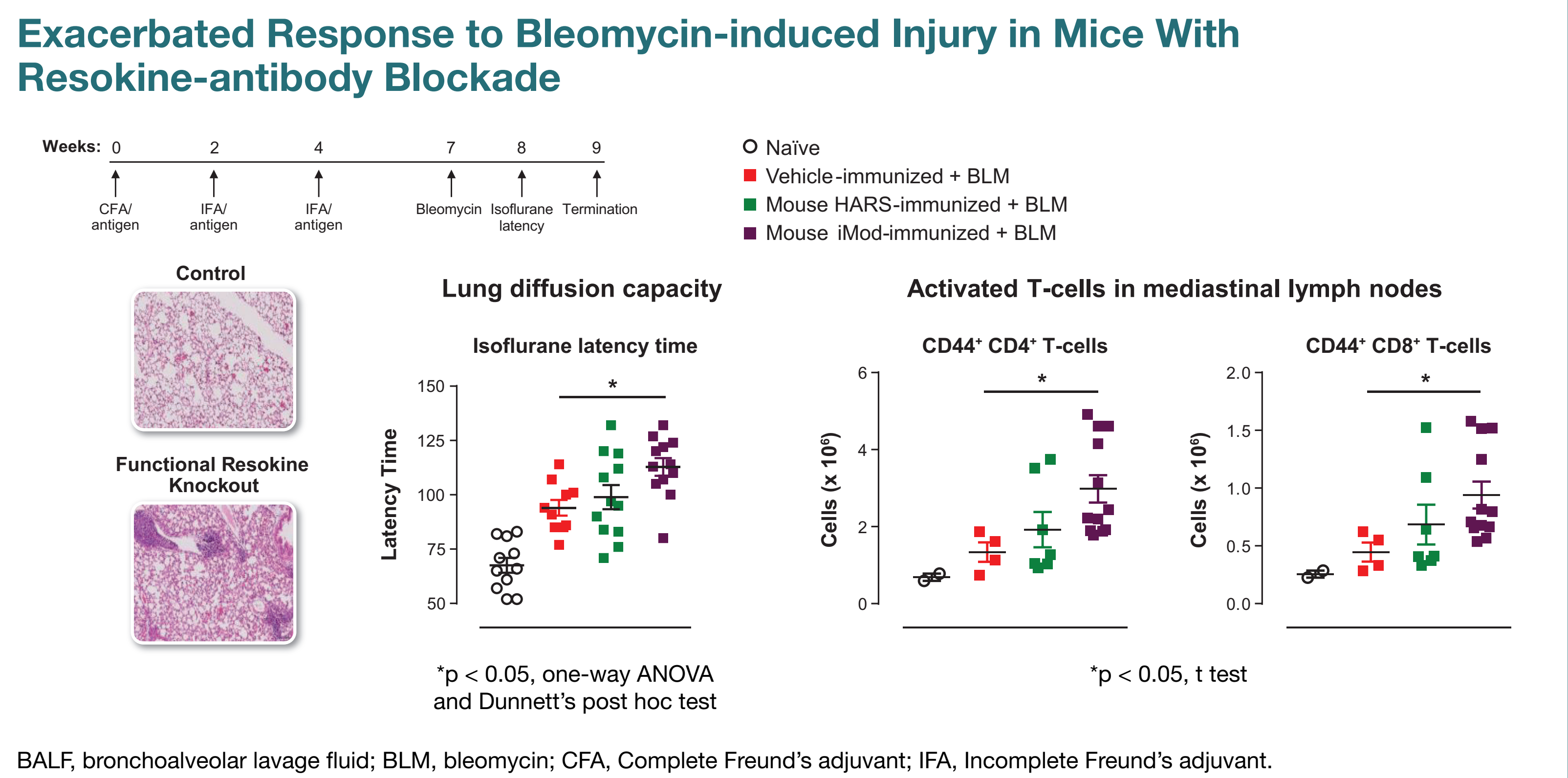
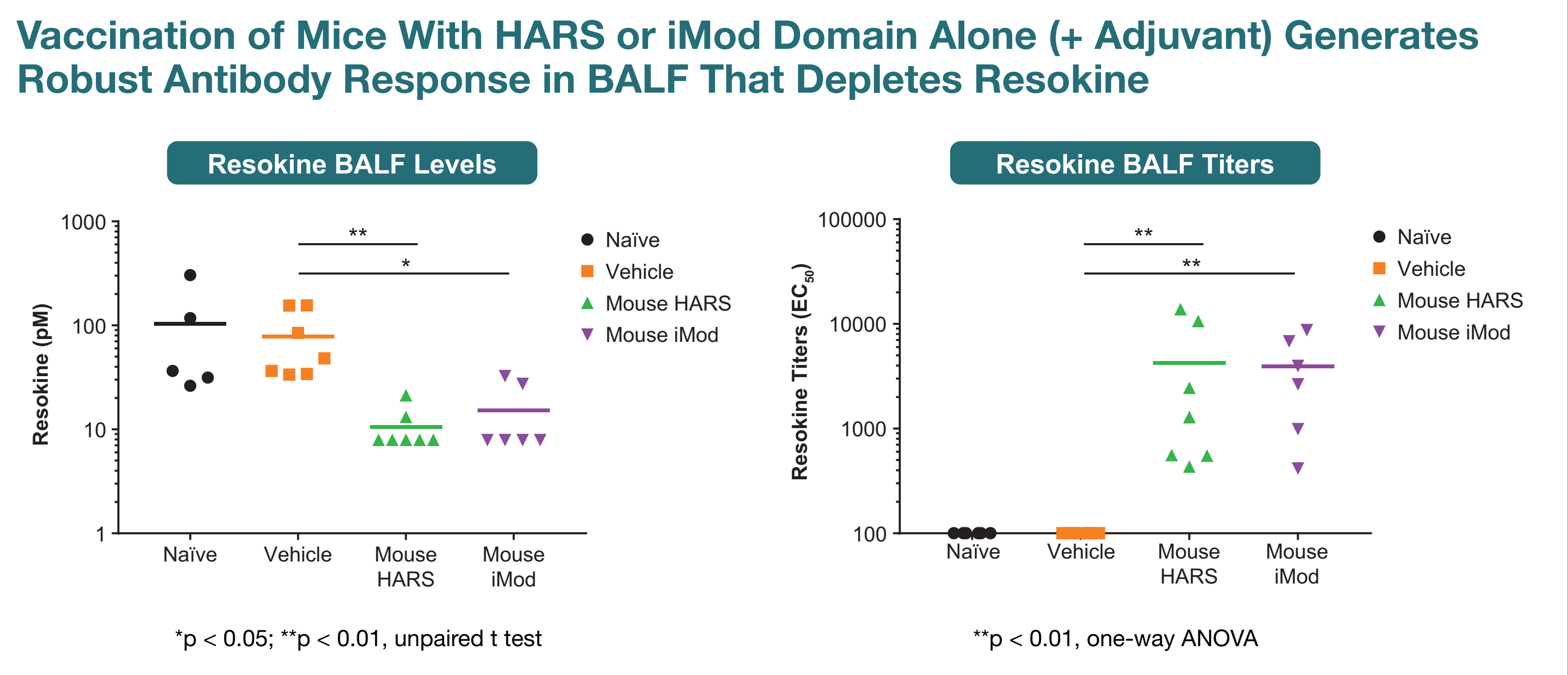
Evolution-elaborated Physiological Systems for Homeostasis



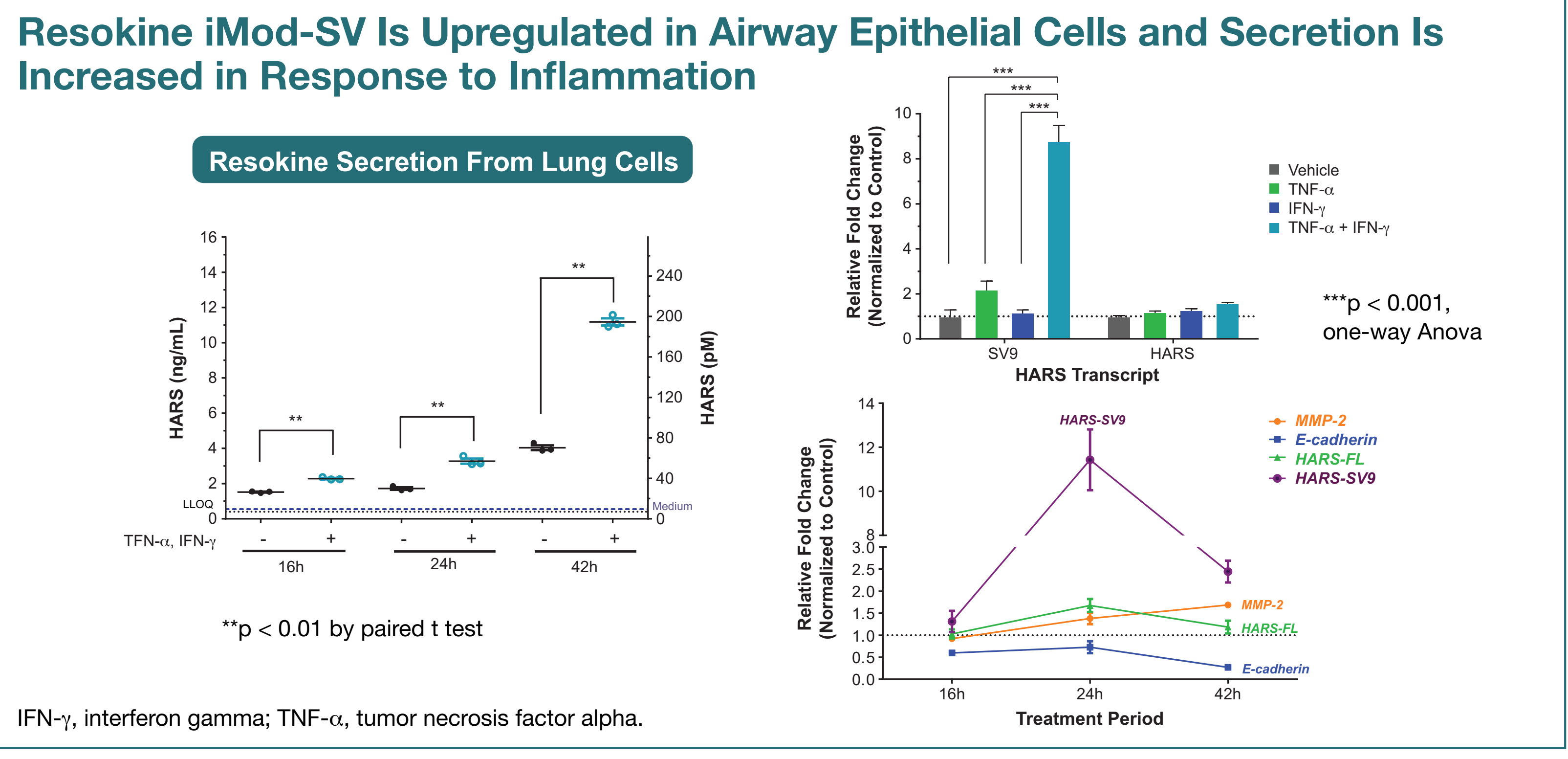
Disrupting the Resokine Pathway Promotes Lung Damage



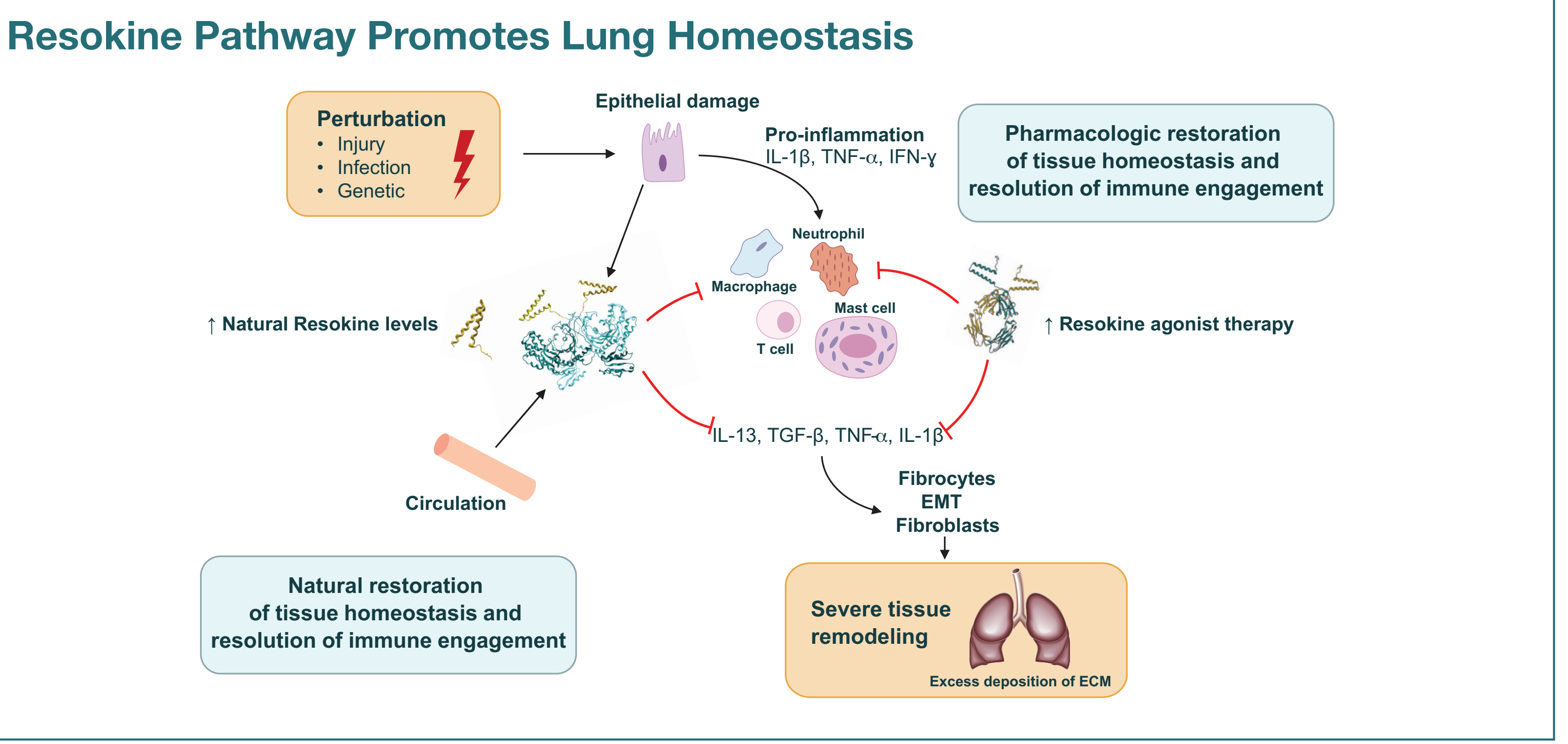
Pathological Remodeling of the Lung in a Model of Resokine Pathway Disruption



Regulation of Resokine Splice Variant in Inflammation



Regulation of the Resokine Pathway in Lung Inflammation



Results

- We characterized serum from a number of Jo-1-positive patients and found the majority exhibited strong cross-reactivity with the N-terminal portion of Resokine and that a splice variant (SV9), which encodes only the N-terminal domain of the protein, is enriched in human lung tissue. Here, we demonstrate that in the lung-derived adenocarcinoma cell line A549, Resokine is actively secreted following stimulation with the inflammatory cytokines IFN-γ and TNF-α, two key players in the initiation of lung inflammation and fibrosis. Secretion is dose dependent and synergistic. SV9, but not the full-length mRNA, is increased following inflammatory cytokine stimulation and its peak expression precedes that of the secreted protein.
- Repeated vaccination of mice with murine Resokine in the presence of adjuvant can break tolerance and generate auto-antibodies. Resokine was present in bronchoalveolar lavage fluid of controls and undetectable in vaccinated animals. In animals with a high titer to Resokine, infiltration of immune cells (including T-cells) into skeletal muscle and lung was detected. Furthermore, in vaccinated animals receiving bleomycin intratracheally, lung function was more severely impacted and both CD4+ and CD8+ T-cells were increased in the mediastinal nodes. Interestingly, these pathologies were observed only in mice that carry a genetic mutation in dysferlin (and other loci) but not in wild-type mouse strains (e.g., C57Bl/6).

Conclusion

- Immune invasion pathologies observed in mice forced to break tolerance to Resokine in the background of another tissue-damaging genetic mutation are similar to activity observed in patients with Jo-1 autoantibodies. This suggests that induction of Resokine insufficiency may disrupt immune homeostasis in a manner that manifests as pathological in lung and muscle. The findings that Resokine circulates at pM levels and the splice variant SV9 is synergistically induced and secreted by lung-derived cells in an inflammatory environment provide evidence for an extracellular niche in which autoantibodies could mediate neutralization of these proteins to induce pathology.

References

Zhou JJ et al. Secreted histidyl-tRNA synthetase splice variants elaborate major epitopes for autoantibodies in inflammatory myositis. *J Biol Chem*, 2014.

Acknowledgements

Alina He (Jo-1 antibody analysis), Kenny D'Arigo/John Bruner (Resokine and Resokine-antibody analysis), Jeanette Ampudia (flow cytometry), Carol Lau (cell culture), Zhiwen Xu, Cario Lo, and Carol Lau (experimental advice, primer design and testing, cell culture). Graphics support was provided by Oxford PharmaGenesis, Inc. and was funded by aTyr Pharma, Inc.

Disclosure

This study was funded by aTyr Pharma, Inc.